

Infant Botulism

Disease Plan

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Last updated: 02/01/2021 by Delaney Moore.

Questions about this disease plan?

Contact the Utah Department of Health Bureau of Epidemiology: 801-538-6191.



CRITICAL CLINICIAN INFORMATION

Clinical Evidence

Signs/Symptoms

- Constipation, poor feeding, decreased sucking, drooling, lethargy, listlessness, ptosis, difficulty swallowing, weak cry, and lack of muscle tone ("floppy baby syndrome")
- · Paralysis and respiratory arrest may occur.

Period of Communicability

• Botulism is not spread person-to-person, but patients with infant botulism can excrete *C. botulinum* toxin and organism in their stool for weeks to months after illness onset.

Incubation Period

2-4 weeks

Mode of Transmission

- Ingestion of food, soil, or dust contaminated with botulinum spores
- Honey is the only food which has been definitively linked to infant botulism and should not be fed to infants <1 year of age.

Laboratory Testing

Type of Lab Test/Timing of Specimen Collection

- Botulism can be diagnosed by culturing the organism itself or by identifying its toxin.
- Botulism testing can only be performed at the Utah Public Health Laboratory and must be approved by the Utah Department of Health Bureau of Epidemiology.
- Test sensitivity decreases when specimen collection is delayed, so specimens should be collected early in the course of illness.

Type of Specimens

Stool or enema

Treatment Recommendations

Type of Treatment

• Botulism Immune Globulin (BabyBIG®) is the preferred treatment and is available through the California Department of Health, Infant Botulism Treatment and Prevention Program (IBTPP).

Time Period to Treat

 BabyBIG® should be administered as soon as possible after symptom onset and should not be delayed until laboratory results are available.

Prophylaxis

None

Contact Management

Isolation of Case

No isolation measures, but meticulous handwashing after changing diapers is required.

Quarantine of Contacts

None

Infection Control Procedures

• In a hospital setting, soiled diapers should be autoclaved and should not be handled by staff with open lesions.

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WHY IS INFANT BOTULISM IMPORTANT TO PUBLIC HEALTH?

Infant botulism results when an infant (usually between the ages of 6 weeks to 6 months) ingests botulism spores which colonize the gastrointestinal tract and produce botulism toxin. Although the illness is rare (on average only 2-4 cases are reported each year in Utah), it is potentially life-threatening and often results in prolonged hospitalization and lengthy rehabilitation. Timely treatment with Botulism Immune Globulin (Baby BIG®) is critical in reducing duration of hospitalization and improving the outcomes of affected infants. In addition, investigation of cases can identify possible clusters and novel exposures (such as contaminated baby food). Public health investigators also play an important role in educating parents of affected infants about precautions that should be taken when the infant comes home from the hospital to prevent other household members and/or pets from being exposed to botulism toxin.



DISEASE AND EPIDEMIOLOGY

Clinical Description

Infant (intestinal) botulism has a distinctly different clinical presentation than wound or foodborne botulism. In infant botulism, the *C. botulinum* spores are ingested, and the toxin is formed in the intestines. It is a rare disease, confined exclusively to infants <1 year of age. The earliest clinical sign in infants is constipation, which is followed by poor feeding, decreased sucking, drooling, lethargy, listlessness, ptosis (drooping eyelids), difficulty swallowing, a weak cry, and lack of muscle tone ("floppy baby syndrome"). Paralysis is generally symmetric and descending. In some cases, respiratory insufficiency and respiratory arrest may occur. Infant botulism presents with a wide range of severity, from mild illness to sudden death. Some studies suggest that infant botulism may be responsible for up to 5% of cases of Sudden Infant Death Syndrome (SIDS).

Causative Agent

Botulism is caused by a potent neurotoxin produced by *Clostridium botulinum*, an anaerobic, spore-forming bacterium. While the bacterium itself is harmless, *C. botulinum* toxin is one of the most potent, lethal substances known. There are seven types of botulinum toxins (A–G), but infant botulism is primarily caused by types A and B. Most cases of infant botulism in Utah are type A, which is more prevalent in the Western United States. Other novel *Clostridia* strains, including *C. baratti* and *C. butyricum*, can also produce a botulinum-like toxin and cause infant botulism.

Differential Diagnosis

There are many conditions and diseases that have presentations similar to infant botulism. Infants with botulism are often diagnosed with dehydration, sepsis, or meningoencephalitis after presentation of lethargy and irritability. Other conditions which should be considered in the differential diagnosis are drug or chemical poisoning, metabolic disorders, spinal muscular atrophy (SMA), Reye's syndrome, congenital myasthenia gravis, poliomyelitis, stroke, and Guillain-Barré Syndrome (although this is not usually seen in children younger than one year of age).

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Laboratory Identification

The diagnosis of botulism can be confirmed by culturing the organism itself or by identifying its toxin. In early cases, diagnosis is more likely made by toxin assay, whereas persons in the later stages of disease are more likely to be culture positive.

Culture

Appropriate specimens include stool or enema.

Toxin Neutralization

Appropriate specimens include stool or enema. It may be possible to identify toxin in serum; however, stool is the preferred specimen. Botulinum toxin in the patient's serum or stool is demonstrated by a toxin neutralization bioassay in mice. This is performed by injecting serum or buffered supernatant from stool into mice and looking for signs of botulism.

Specimen Collection

Fecal specimens should be collected in a sterile container with a tight, screw-capped lid, such as a urine specimen container. Do not use containers with fixatives, such as those used for ova and parasite collection. If no stool can be collected spontaneously or through gentle digital stimulation, an enema should be collected. However, **glycerin suppositories yield an unsatisfactory specimen and should not be used**.

Testing Protocol

Botulism testing is time, labor, and resource intensive. Unlike other laboratory tests, the test for botulism is not generally used as a rule-out test. While botulism testing is highly specific, sensitivity is quite low. This means that a positive test can be interpreted as positive in almost all cases, but a negative test is not conclusive. Test sensitivity is decreased when specimen collection is delayed. The lead enteric epidemiologist or medical officer at the Utah Department of Health (UDOH) Bureau of Epidemiology (BOE) reviews all botulism test requests. Test requests are approved after consultation with the requesting physician and the Utah Public Health Laboratory (UPHL).

The Utah Public Health Laboratory is the only laboratory in Utah that offers botulism testing. Both culture and toxin neutralization tests are available.

Treatment

Most infant botulism patients require hospitalization. Botulism Immune Globulin (BabyBIG®) is effective in reducing the length of hospital stay and cost for infant botulism patients. It is available through the California Department of Health, Infant Botulism Treatment and Prevention Program (IBTPP). Prompt treatment with BabyBIG® immediately ends toxemia and enables nerve regeneration to begin. The treatment is most effective when provided as soon as possible after symptom onset. Treatment should not be delayed until laboratory results are available. Physicians requesting BabyBIG® should contact the IBTPP on-call physician directly at (510) 231-7600 (available 24 hours/day). More information is available on the IBTPP website at http://www.infantbotulism.org/. Requesting BabyBIG® does not replace reporting requirements to public health.

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Case Fatality

Among hospitalized cases in the U.S., the case fatality rate for infant botulism is <1%.

Reservoir

Clostridium botulinum spores are ubiquitous in the environment. The spores can survive indefinitely in soil under almost any environmental condition and can travel through dust. Although botulism spores can be found in a variety of foods, honey is the only food which has been definitively linked to development of infant botulism.

Transmission

Infant botulism occurs when *C. botulinum* spores germinate and produce toxin in the anaerobic conditions of the gastrointestinal tract of infants. This can happen through ingestion of food, soil, or dust contaminated with botulinum spores. This kind of infection is rare in adults because the natural bacterial flora in adult gastrointestinal tracts inhibit the germination of *C. botulinum* spores, and thus the production of botulinum toxin. However, *C. botulinum* spores can germinate and produce toxin in intestines of adults whose natural flora have been disrupted by disease or antibiotic use. Honey often contains *C. botulinum* spores. Some cases of intestinal botulism have occurred in infants living in areas of construction and earth disruption. The stools of affected infants contain toxin and may pose a risk to household members and pets.

Susceptibility

All infants are potentially susceptible to infant botulism. Both formula-fed and breast-fed infants are at risk; however, onset of illness in formula-fed infants occurs at a significantly younger age than breast-fed infants (7.6 weeks vs. 13.7 weeks). Introduction of solid foods may aid colonization with *C. botulinum* by causing changes in the gut microflora. In addition, infants with slow gut motility (defined as having less than one stool per day) may be at increased risk for developing infant botulism.

Infants treated with BabyBIG® will have a protective level of toxin-neutralizing antibody for at least six months following administration of the medicine, and during this time an infant cannot get botulism again. There have been no reported cases of an infant acquiring botulism more than once. However, in some cases, infants who are prematurely discharged from hospital care may have a relapse of breathing and feeding difficulties.

Incubation Period

The incubation period for infant botulism is 2-4 weeks.

Period of Communicability

While botulism is not spread from person-to-person, patients with infant botulism excrete both *C. botulinum* toxin and organism in their feces from weeks to months after onset. Scrupulous handwashing should be practiced after diaper changes, and persons with cuts or open wounds on their hands should wear gloves when changing diapers. Soiled diapers should be disposed of where no person or animal can come into contact with them.

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Epidemiology

Infant botulism, the most common form of botulism, has only been recognized since 1976. It is a rare disease with an average of 2-4 cases reported each year in Utah. However, Utah has the third highest incidence of infant botulism in the U.S., following Delaware and Hawaii. This may be partly due to increased physician awareness and access to diagnostic testing. Interestingly, eight of 11 states with the highest incidence are located west of the Rocky Mountains.



PUBLIC HEALTH CONTROL MEASURES

Public Health Responsibility

The public health investigator should ensure that the patient has received BabyBIG®. This is typically obtained by the treating physician directly from the IBTPP. If infant botulism is strongly suspected, but the physician is not sure whether treatment is warranted due to delayed diagnosis, consultation with the IBTPP is strongly encouraged. The phone number for the on-call IBTPP physician is 510-231-7600.

The investigator should obtain appropriate hospital records, including Admission Notes and Discharge Summary. The patient's parent or guardian should be interviewed using the Infant Botulism case report form. Education should be provided to all caregivers regarding appropriate handing of diapers.

Prevention

The only known prevention measure for infant botulism is to avoid feeding honey to infants 12 months of age or less. Breastfeeding may slow the onset and lessen the severity of illness.

Chemoprophylaxis

None.

Vaccine

None.

Isolation and Quarantine Requirements

Isolation: Isolation measures are not required, but meticulous handwashing is required. Because the patient may excrete the toxin and organism for weeks to months, close contact with other infants and children should be limited during this time to ensure that other children do not come into contact with fecal material from a leaky diaper. Any contact the patient has with other infants and children during this time should be supervised by an adult.

C. botulinum is not part of the patient's normal flora and will eventually stop being excreted in the infant's feces.

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Hospital: In the hospital, soiled diapers should be autoclaved and should not be handled by staff with open lesions.

Quarantine: None.



CASE INVESTIGATION

Reporting

Infant botulism cases are not immediately notifiable. Cases should be reported within three working days after identification in Utah.

Table 1: Criteria to determine whether a case should be reported

*Criteria which apply to infant botulism

Criterion			Reporting	
Historical Evidence				
History of a fresh, contaminated wound during the two weeks before onset of			0	
symptoms				
Ingestion of home-canned food within the 48 hours before onset of symptoms			0	
Clinical Evidence				
Diplopia (double vision)		Ο	0	
Blurred vision		0	0	
Bulbar weakness		0	0	
Impaired respiration [±]		O±	O±	
Progressive weakness [±]		O±	O±	
Progressive symmetric paralysis		0	0	
Healthcare record contains a diagnosis of botulism [±]	S±			
Death certificate lists botulism as a cause of death or a significant condition	S±			
contributing to death [±]	3			
Laboratory Evidence				
Detection of botulinum toxin in serum	S*			
Detection of botulinum toxin in stool	S*±			
Detection of botulinum toxin in patient's food	S*			
Isolation of Clostridium botulinum from stool	S*±			
Isolation of Clostridium botulinum from wound	S*			
Epidemiologic Evidence				
Ingestion of the same food as persons who have laboratory-confirmed botulism		Ν		
Special Criteria				
Request for anti-toxin specific for botulinum toxin	S*			
Request for BabyBIG from the Infant Botulism Treatment and Prevention Program [±]	S*±			

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Notes:

S = This criterion alone is Sufficient to identify a case for reporting.

N = All "N" criteria in the same column are Necessary to identify a case for reporting.

O = At least one of these "O" (Optional) criteria in each category (i.e., clinical evidence and laboratory evidence) in the same column—in conjunction with all "N" criteria in the same column—is required to identify a case for reporting. (These optional criteria are alternatives, which means that a single column will have either no O criteria or multiple O criteria; no column should have only one O.)

*A requisition or order for any of the "S" laboratory tests is sufficient to meet the reporting criteria.

Case Definition

Clinical description

An illness of infants, characterized by constipation, poor feeding, and "failure to thrive" that may be followed by progressive weakness, impaired respiration, and death.

Laboratory criteria

- Detection of botulinum toxin in stool or serum, OR
- Isolation of Clostridium botulinum from stool.

Case classification

Confirmed: A clinically compatible case that is laboratory-confirmed, occurring in a child aged less than one year.

Table 2: Criteria for defining a case of botulism

*Criteria which apply to infant botulism

Criterion		Reporting		
Historical Evidence		Confirmed		
History of a fresh, contaminated wound during the two weeks before onset of symptoms	N3,A4			
Ingestion of home-canned food within the 48 hours before onset of symptoms	A3,A4		N1	
Clinical Evidence				
Diplopia (double vision)	01,03,04	01	O1,O3,O	
Blurred vision	01,03,04	01	O1,O3,O	
Bulbar weakness	01,03,04	01	O1,O3,O	
Impaired respiration [±]	O2±		O2±	
Progressive weakness [±]	O2±		O2 [±]	
Progressive symmetric paralysis	01,03,04	01	O1,O3,O	
Healthcare record contains a diagnosis of botulism				
Death certificate lists botulism as a cause of death or a significant condition contributing to death				
Laboratory Evidence				
Detection of botulinum toxin in serum	0	0		
Detection of botulinum toxin in stool [±]	O1,O2 [±] ,O4	01		

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Detection of botulinum toxin in patient's food	01,04	01	
Isolation of Clostridium botulinum from stool±	O1,O2±,O4		
Isolation of Clostridium botulinum from wound	O3,O4		
Epidemiologic Evidence			
Ingestion of the same food as persons who have laboratory-confirmed botulism	A3,A4	01	A?
Special Criteria			
Age <1 year	N2±,A4±		

Notes:

N = All "N" criteria in the same column are Necessary to classify a case.

A = This criterion must be absent (i.e., NOT present) for the case to meet the classification criteria.

O = At least one of these "O" (Optional) criteria in each category (i.e., clinical evidence and laboratory evidence) in the same column—in conjunction with all "N" criteria in the same column—is required to classify a case.

- 1 = Foodborne botulism
- 2 = Infant botulism[±]
- 3 = Wound botulism
- 4 = Other botulism

Case Investigation Process

In most cases, identification of the source of infection is not possible. This is because *C. botulinum* spores are ubiquitous in the soil and dust. Honey has been identified as a vehicle of *C. botulinum* spores. However, investigators should keep in mind less common or less frequently identified sources of infant botulism. Investigators should also be vigilant of foodborne botulism cases in an infant that may present as infant botulism. Botulism toxin has been identified in home canned baby foods.

Public health has three main roles when a case of infant botulism is identified or suspected:

1) Laboratory testing

UPHL is the only laboratory in Utah that provides testing of human and non-human samples for botulinum toxin and *C. botulinum*. The Bureau of Epidemiology (BOE) screens all botulism test requests. The BOE approves tests requests after consultation with the physician and UPHL.

2) Treatment

Healthcare providers work directly with the IBTPP to obtain BabyBIG®. The BOE ensures that this consultation has been initiated when appropriate.

3) Education

The public health investigator should provide appropriate education to the patient's family. The IBTPP website (http://www.infantbotulism.org/) is an excellent resource.

Outbreaks

Outbreaks of infant botulism have not been reported. However, geographic clustering of cases has been seen, and would warrant investigation and reporting to the Centers for Disease Control and Prevention (CDC).

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Identifying Case Contacts

Adult contacts of a case of infant botulism are not at risk of developing disease.

Case Contact Management

Not applicable.

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VERSION CONTROL

Updated February 2016: Created new disease plan specific to infant botulism.

Updated February 2021: Added Critical Clinician Information and Electronic Laboratory Reporting Processing Rules sections. All other sections updated.

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UT-NEDSS (EpiTrax) Minimum/Required Fields by Tab

Demographic

- First Name
- Last Name
- Street Number
- Street Name
- City
- State
- County
- Zip Code
- Date of Birth
- Area Code
- Phone Number
- Birth Gender
- Ethnicity
- Race

Clinical

- Disease
- Onset Date
- Visit Type
 - (if inpatient) Did Botulism cause hospitalization?
- Died
 - o (if yes) Date of Death
 - o (if yes) Did Botulism cause death?
- Was BabyBIG administered?
 - o (if yes) Date administered
- Diagnostic Facility
- Symptoms

Laboratory

- Lab Name
- Lab Test Date
- Collection Date
- Specimen Source
- Test Type

- Organism
- Test Result
- Accession Number
- Was specimen or isolate forwarded to CDC for testing or confirmation?
 - o (if yes) Date specimen sent to CDC

Epidemiological

- Childcare Association
 - Name of the childcare
 - Location
 - O Did the patient work/attend while ill?
 - Important information including dates
- Imported From
- Risk Factors
- Risk Factor Notes

Investigation

- Date 30 days before disease onset
- Date 3 days before disease onset

Contacts

- Does case's infection appear secondary to another person's infection? (if YES, please fill out info in contact table)
- Any contacts ill with similar symptoms? (if YES, please fill out info in contact table)

Reporting

• Date first reported to public health

Administrative

- State Case Status
- Outbreak Associated
- Outbreak Name

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ELECTRONIC LABORATORY REPORTING PROCESSING RULES

Infant Botulism Rules for Entering Laboratory Test Results

The following rules describe how laboratory results reported to public health should be added to new or existing events in UT-NEDSS (EpiTrax). These rules have been developed for the automated processing of electronic laboratory reports, although they apply to manual data entry, as well.

Test-Specific Rules

Test specific rules describe what test type and test result combinations are allowed to create new morbidity events in UT-NEDSS (EpiTrax), and what test type and test result combinations are allowed to update existing events (morbidity or contact) in UT-NEDSS (EpiTrax).

Test Type	Test Result	Create a New Event	Update an Existing Event
PCR/Amplification	Positive	Yes	Yes
	Negative	No	Yes
PCR/Amplification	Equivocal	No	Yes
	Other	No	Yes
	Positive	Yes	Yes
Culture	Negative	No	Yes
Culture	Equivocal	No	Yes
	Other	No	Yes

Whitelist Rules

Whitelist rules describe how long an existing event can have new laboratory data appended to it. If a laboratory result falls outside the whitelist rules for an existing event, it should not be added to that event, and should be evaluated to determine if a new event (CMR) should be created.

Infant Botulism Morbidity Whitelist Rule: If the specimen collection date of the laboratory result is two years or less after the last positive specimen collection date of the morbidity event, the laboratory result should be added to the morbidity event.

Infant Botulism Contact Whitelist Rule: Never added to contact.

Graylist Rule

We often receive laboratory results through ELR that cannot create cases, but can be useful if a case is created in the future. These laboratory results go to the graylist. The graylist rule describes how long an existing event can have an old laboratory result appended to it.

Infant Botulism Graylist Rule: If the specimen collection date of the laboratory result is 30 days before to seven days after the event date of the morbidity event, the laboratory result should be added to the morbidity event.

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Other Electronic Laboratory Processing Rules

• If an existing event has a state case status of "not a case," ELR will never add additional test results to that case. New labs will be evaluated to determine if a new CMR should be created.

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